

Beverly Wendland, Ph.D.

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Executive Profile

Transformational academic leader, champion of basic and applied research to solve societal challenges, and organization builder, having established 8 institutes, centers, and schools over the past decade. Served as Provost of Washington University in St. Louis and Dean of the Krieger School of Arts and Sciences at Johns Hopkins University, leading both institutions through critical junctures by developing strategic roadmaps, restructuring governance, recruiting marquee faculty talent, and securing more than \$800M in philanthropic support. First-generation college student able to connect with all audiences from undergraduates to trustees and community members to business leaders.

Leadership Experience

Washington University in St. Louis

Provost and Executive Vice Chancellor for Academic Affairs | July 2020 – July 2025

Senior Advisor to the Chancellor | As of August 2025

Appointed the university's second-in-command, guided academic affairs for nine schools on the Danforth Campus, and supplied institutional leadership across undergraduate, graduate, and professional programs. Oversaw a \$900M budget and eight schools with 15,532 undergraduate, graduate, professional students, 1639 faculty, and 1500+ staff. As the transformational force behind WashU's new School of Public Health, owned the full academic lifecycle, from program design to faculty affairs, recruiting, teaching, research, and student experience. Charted a bold roadmap that reimaged academic infrastructure, setting a new standard for scalable, mission-aligned operations in higher education.

- Mobilized \$1B in unrestricted endowment returns to advance transformation, allocating \$800M to eliminate undergraduate loans and implement need-blind admissions, \$200M to expand graduate fellowships, and \$350M to fund cross-cutting academic priorities through the Provost's strategic initiative portfolio.
- Unified institutional strategic planning across faculty and trustees through *Here and Next*, winning Trustee support by inspiring research and improving Danforth's partnership with Washington University's world-class School of Medicine.
- Established the university's new School of Public Health with an innovative and interdisciplinary ethos, anchoring the #17 ranked MPH program. Secured \$8M in philanthropic gifts to endow a deanship and professorship, recruited preeminent scholar Sandro Galea as founding dean, and realized an 87% spike in applicant volume.
- Converted a trustee's insight into a \$50M gift to launch the Food and Agriculture Research Mission (FARM), an interdisciplinary initiative housed in the School of Public Health addressing food insecurity and promoting sustainable agriculture.

- Recruited 15 nationally recognized “Field Leading Faculty,” increasing the university’s profile as a research powerhouse, and stimulating interdisciplinary research through the *Here and Next* Seed Grant Program. Turned \$4.6M in initial funding into \$33.2M in external awards, with an additional \$35.1M pending. Fostered interdisciplinary scholarship through initiatives such as EnviroTex and Trusted Tap, united faculty from engineering, arts and sciences, public health, and design in pursuit of practical solutions to public challenges. Grew external funding by 42% over three years.
- Executive sponsor of the \$125M *Student Sunrise* student information system technology transformation project. Delivered this pivotal uplift on time and \$5M under budget, restoring campus trust after a previous challenging rollout of Workday HR/FIN and positioning WashU as a Workday Student implementation benchmark across higher education.
- Catalyzed planning and institutional support behind Missouri’s first NSF-funded engineering and research center (CURB), a \$26M grant, to advance carbon utilization redesign and biomanufacturing-empowered decarbonization.
- Redesigned academic governance and institutional accountability by restructuring graduate education under new Vice Provost for Graduate Education and Doctoral Council, building the Office of Institutional Effectiveness to support data-informed decision-making, and deepening trustee engagement through redesigned board reporting and strategic communications.

Johns Hopkins University

James B. Knapp Dean, Krieger School of Arts and Sciences | 2015 – 2020

Interim Dean | 2014 – 2015

Served as chief academic and administrative officer of JHU’s most diverse academic division, overseeing 22 departments and multiple interdisciplinary centers and institutes. Oversaw a \$426M Budget with 8,252 undergraduate, masters, Ph.D. students, 610 faculty and 457 staff. Guided end-to-end academic programs, faculty hiring and development, research growth, and operational performance, while cultivating philanthropic partnerships and deepening trustee and alumni engagement through the Krieger School Advisory Board.

- Raised \$747M against a \$550M fundraising goal. Founded the Alexander Grass Humanities Institute (\$10M gift) and the SNF Agora Institute (\$150M gift). Strengthened civic engagement, scholarship, and interdisciplinary collaboration. Secured naming gift for Miller Department of Philosophy (\$75M gift), the largest ever to a Philosophy Department.
- Expanded continuing education through Advanced Academic Programs by doubling number of masters programs and certificates offered to 26 total and tripling gross revenue to \$80M. New STEM and business offerings focused on fulfilling workforce development opportunities.
- Transformed graduate education governance by building guardrails, including centralized tracking, compliance, and student support systems, replacing a decentralized model. Added structured advising and clear expectations with defined milestones.
- Maintained USNWR Top 10 ranking (2015-2020), moving up to #9 in 2020. Prioritized operational excellence by advancing the *Ten by Twenty* strategic plan and focusing on high-impact academic discourse and scholarship.

Academic Appointments

Washington University in St. Louis

- Provost and Executive Vice Chancellor for Academic Affairs, 2020–2025
- Professor of Biology (tenured), 2020–Present

Johns Hopkins University

- Dean, Krieger School of Arts and Sciences, 2015–2020
 - Interim Dean, Krieger School of Arts and Sciences, 2014–2015
 - Chair, Department of Biology, 2009–2014
 - Co-Director, CMDB Graduate Program, 2006–2009
 - Professor of Biology (tenured), 2008–2020
 - Associate Professor of Biology, 2004–2008
 - Assistant Professor of Biology, 1998–2004
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Education

- Ph.D., Neurosciences, Stanford University; Stanford, CA
Thesis: *Molecular mechanisms of regulated exocytosis*
 - B.S., Bioengineering, magna cum laude, University of California, San Diego; La Jolla, CA
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Honors & Recognition

- America's Power List, recognized for excellence (2024)
 - Fellow, American Association for the Advancement of Science (2015)
 - Multiple national honors for mentoring, advising, and public service
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Professional Engagement & Service

- Member, Barnes Jewish Hospital Board of Directors, St. Louis, MO (2020–2025)
- Member, Springer Nature US Research Advisory Board (2025–present)
- Editorial boards: Prior service on *Molecular Biology of the Cell*, *EMBO Reports*, *Traffic*, among others
- NIH and NSF grant and program reviewer; served on multiple national advisory panels
- Organizer and speaker for numerous scientific meetings and public policy forums
- Member: AAAS, ASCB

Research & Scholarship

Selected Grant Funding

- NSF ADVANCE IT grant, co-PI: *AIM for Equity* (2024–2025)
- NSF, NIH R01 and T32 grants supporting research and graduate training in molecular cell biology
- PI or co-PI on numerous awards totaling millions in federal and foundation funding

Meeting Presentations

2024

- ASCB Annual Meeting, WICB Panel and Networking Session: Academic Evolution: Rethinking Metrics, Rewriting Rules, San Diego, CA
- Symposium to celebrate the retirement of Dr. Scott Emr, La Jolla, CA

2023

- AAU Annual Provost Conference, panelist on “The Impact of Artificial Intelligence on Higher Education.” AAU Headquarters, Washington, D.C.

2014

- Lysosomes and Endocytosis Gordon Conference. Proctor Academy, Andover, NH., Discussion Leader for Keynote Session and meeting Chair/organizer.
- Symposium on Cell Signaling and Membrane Traffic, La Jolla, CA
- FASEB Meeting on Methods in Protein Structure Analysis, Snowmass, CO
- Second Annual BMCB & GGD Symposium, Cornell University, Ithaca, NY (student organized symposium)

2013

- Synaptic Vesicle Biogenesis, HHMI/Janelia Farm Conference. Ashburn, VA
- ASCB Annual Meeting, Symposium Speaker, New Orleans, LA

2012

- Biophysical Society Annual Meeting; Exo-Endocytosis Subgroup Speaker. San Diego, CA
- Lysosomes and Endocytosis Gordon Conference. Proctor Academy, Andover, NH., Discussion Leader and meeting co-Chair/co-organizer.

2011

- ASM CUE, Learn Something New session. Baltimore, MD
- Molecular Membrane Biology Gordon Research Conference, Session Chair. Andover, NH

2010

- Keystone Symposium, Molecular Basis for Biological Membrane Organization and Dynamics, Jan 10 - 15, 2010, Snowbird, Utah., Invited Speaker
- Lysosomes Gordon Research Conference, Session Chair. Andover, NH

2008

- Plant and Fungal Cytoskeleton Gordon Conference. Il Ciocco, Lucca, Italy. Invited Speaker.

2007

- Yeast Cell Biology meeting, Membrane Trafficking session. Cold Spring Harbor Labs. Cold Spring Harbor, NY. Session Chair and speaker.

2004

- Lysosomes and Endocytosis Gordon Conference. Proctor Academy, Andover, NH., Session Chair.

2003

- 33rd Annual Society for Neuroscience Meeting. New Orleans, LA. Invited Symposium speaker.

2002

- Cellular and Molecular Mycology Gordon Conference. Holderness School, Plymouth, NH., Invited Speaker.
- Lysosomes Gordon Conference. Proctor Academy, Andover, NH., selected abstract.

2001

- Phosphoinositides and the Golgi. NIH, Bethesda, MD., Invited Speaker.
- Keith R. Porter Symposium on Cytoplasmic Organization and Membrane Traffic. Airlie House, Airlie, VA., selected abstract.
- Endocytosis Minisymposium. 41st Annual Meeting of the American Society for Cell Biology. Washington, D.C., Invited Session Co-chair and speaker.

2000

- Endocytosis and Actin session. 3rd Annual JHU/NIH Membrane Trafficking Workshop. Annapolis, MD., Invited Speaker.
- Lysosomes Gordon Conference. Proctor Academy, Andover, NH., selected abstract.

1999

- Endocytosis Minisymposium 39th Annual Meeting of the American Society for Cell Biology. Washington, D.C., selected abstract.

1997

- Special Interest Group on Molecular Mechanisms of Vesicle Formation. 37th Annual Meeting of the American Society for Cell Biology. Washington, D.C., Invited Speaker

Invited Seminars

2014

- Johns Hopkins University Bloomberg School of Public Health, Department of Biochemistry and Molecular Biology. Baltimore, MD

2013

- University of Pittsburgh School of Medicine, Department of Cell Biology. Pittsburgh, PA.
University of Pennsylvania School of Medicine, Children's Hospital of Philadelphia, Division of Cell Pathology. Philadelphia, PA.

2012

- Duke University, Program in Genetics and Genomics. Durham, NC 2011
- Dartmouth College School of Medicine, Department of Biochemistry. Keynote speaker at departmental retreat. Morey Lake, NH.

2010

- University of Massachusetts Medical School, Department of Biochemistry and Molecular Pharmacology, Worcester, MA
- Johns Hopkins University, Department of Biological Chemistry, Baltimore, MD
- University of Nebraska Medical Center, Invited by the graduate students from the Department of Biochemistry and Molecular Biology, Omaha, NE
- Johns Hopkins University, Department of Biology, Colloquium Speaker. Baltimore, MD

2008

- University of Toronto, Cell Biology Program, Toronto, Ontario, Canada

2007

- University of Washington, Dept. of Biochemistry, Seattle, WA
- University of British Columbia, Centre for Mol. Med. and Therap., Vancouver, BC, Canada
- Killiam Seminar Series, McGill Univ., Montreal Neurological Inst., Montreal, Quebec, Canada
- Northwestern University, Dept. of Biochem., Mol. Biol. and Cell Biol., Evanston, IL

2006

- University of Puerto Rico, Rio Piedras, Dept. of Biology, San Juan, Puerto Rico
- University of Iowa, Dept. of Physiology and Biophysics, Iowa City, IA
- University of Utah, Dept. of Biology, Salt Lake City, UT

2005

- Weill Medical College of Cornell University, Dept. of Biochemistry, New York, NY
- Queen's College, Dept. of Biology, Flushing, NY
- Johns Hopkins Medical School, Dept. of Cell Biology, Baltimore, MD

2004

- University of Pennsylvania, Dept. of Cell and Developmental Biology, Philadelphia, PA

2003

- Catholic University of America, Department of Biology, Washington, D.C.
- University of California at Los Angeles School of Medicine, Department of Human Genetics, Los Angeles, CA.
- Yale University School of Medicine, Department of Cell Biology, New Haven, CT.
- University of California at Davis, Joint Seminars in Molecular Biology, Davis, CA.
- Georgetown University, Dept. of Biology, Washington, DC

2002

- University of Texas at Austin, Institute for Cell and Molecular Biology; Austin, TX
- University of Texas Health Science Center, Department of Biochemistry; San Antonio, TX

- Uniformed Services Health Sciences University, Department of Biochemistry; Bethesda, MD
- University of North Carolina, Department of Pharmacology, Raleigh, NC
- National Institutes of Health, Laboratory of Cell Biology, Bethesda, MD
- Stanford University, Department of Molecular and Cellular Physiology, Stanford, CA.
- University of California at Berkeley, Dept. of Mol., Cell and Devel. Biology, Berkeley, CA.
- 2001
- Johns Hopkins University, Department of Biology Colloquium; Baltimore, MD.
- Mayo Clinic, Department of Biochemistry and Cell Biology; Rochester, MN

2000

- Johns Hopkins School of Public Health, Department of Biochemistry; Baltimore, MD.
- University of Maryland College Park, Department of Biology; College Park, MD.
- University of Wisconsin-Madison, Department of Pharmacology; Madison, WI.
- Carnegie Institute of Washington, Department of Embryology; Baltimore, MD.
- Vanderbilt University, Department of Biological Sciences; Nashville, TN.
- University of Alberta, Department of Cell Biology; Edmonton, Alberta, Canada.

1999

- University of Rome, Department of Biology; Rome, Italy
- National Institutes of Health, Laboratory for Cell Biology; Bethesda, MD.
- Johns Hopkins School of Medicine, Department of Anatomy and Cell Biology; Baltimore, MD.

1998

- Johns Hopkins School of Medicine, Department of Neurosciences; Baltimore, MD.

1997

- National Institutes of Health, Laboratory of Cellular and Molecular Biophysics; Bethesda, MD.

Publications

76. Hoban, K., Lux, S.Y., Poprawski, J., Zhang, Y., Shepherdson, J., Castiñeira, P.G., Pesari, S., Prosser, D.C., Norris, C., Wendland, B. ESCRT-dependent protein sorting is required for the viability of yeast clathrin-mediated endocytosis mutants. (2020). *Traffic*. **21**(6):430-450.
75. Wrasman, K., Alioto, SL, Zhang, Y., Hoban, K., Khairy, M., Goode, BL, and **Wendland, B.** A flow cytometry-based phenotypic screen to identify novel endocytic factors in *Saccharomyces cerevisiae*. (2018). *G3*. **8**(5):1497-1512.
74. Apel AR, Hoban K, Chuartzman S, Tonikian R, Sidhu S, Schuldiner M, **Wendland B**, Prosser D. Syp1 regulates the clathrin-mediated and clathrin-independent endocytosis of multiple cargo proteins through a novel sorting motif. (2017). *Mol Biol Cell*. **28**(18):2434- 2448.
73. Prosser DC, Wrasman K, Woodard TK, O'Donnell AF, **Wendland B.** Applications of pHluorin for Quantitative, Kinetic and High-throughput Analysis of Endocytosis in Budding Yeast. (2016). *J Vis Exp*. 2016 Oct 23;(116).
72. Prosser DC, **Wendland B.** DePFth Perception in Clathrin-Mediated Endocytosis. (2016). *Dev Cell*. **37**(5):387-8.

71. Feliziani C, Valdez Taubas J, Moyano S, Quassollo G, Poprawski JE, **Wendland B**, Touz MC. Vestiges of Ent3p/Ent5p function in the giardial epsin homolog. (2016). *Biochim Biophys Acta*. **1863**(4):749-59.
70. Whitfield ST, Burston HE, Bean BD, Raghuram N, Maldonado-Báez L, Davey M, **Wendland B**, Conibear E. The alternate AP-1 adaptor subunit Apm2 interacts with the Mil1 regulatory protein and confers differential cargo sorting. (2016). *Mol Biol Cell*. **27**(3):588-98.
69. Prosser DC, Pannunzio AE, Brodsky JL, Thorner J, **Wendland B**, O'Donnell AF. Alpha-arrestins participate in cargo selection for both clathrin-independent and clathrin-mediated endocytosis. (2015). *J Cell Sci*. 128(22):4220-34.
68. Bradford, M.K., Whitworth, K. and **Wendland, B.** (2015). Pan1 regulates transitions between stages of clathrin-mediated endocytosis. *Mol. Biol. Cell*. **26**:1371-1385.
67. Messa M, Fernandez-Busnadiego R, Sun EW, Chen H, Czapla H, Wrasman K, Wu Y, Ko G, Ross T, **Wendland B**, and De Camilli P. (2014). Epsin deficiency impairs endocytosis by stalling the actin-dependent invagination of endocytic clathrin coated pits. *eLife* **3**:e03311.
66. Lang MJ, Martinez-Marquez JY, Prosser DC, Ganser LR, Buelto D, **Wendland B**, Duncan MC. (2014). Glucose starvation inhibits autophagy via vacuolar hydrolysis and induces plasma membrane internalization by down regulating recycling. *J. Biol. Chem*. **289**:16736-47.
65. Alvaro C.G.*, O'Donnell A.F.*, Prosser D.C.*, Augustine A.A., Goldman A., Brodsky J., Cyert M.S., **Wendland B.**, Thorner J. (2014). Specific α -arrestins negatively regulate *Saccharomyces cerevisiae* pheromone response by down-modulating the G-protein coupled receptor Ste2. *Mol. Cell. Biol*. **34**:2660-2681. * Authors contributed equally
64. Pierce, B.D., Toptygin, D. and **Wendland, B.** (2013). Pan1 is an intrinsically disordered protein with homotypic interactions. *Proteins*. **81**:1944-63.
63. Whitworth, K., Bradford, M.K., Camara, N. and **Wendland, B.** (2013). Targeted disruption of an EH-domain protein endocytic complex, Pan1-End3. *Traffic*. **15**:43-59.
62. B.L. Goode, Eskin J.A., and **B. Wendland**. (2015). Actin and endocytosis in budding yeast. *Genetics*. **199**:315-358.
61. Prosser, D.C. and **B. Wendland**. (2012). Conserved roles for yeast Rho1 and mammalian RhoA GTPases in clathrin-independent endocytosis. *Small GTPases*. 3:229-235.
60. Umasaker, P.K., Sanker, S., Thieman, J.R., Chakraborty, S., **Wendland, B.**, Tsang, M., Traub, L.M. (2012). Distinct and separable activities of the endocytic clathrin-coat components Fcho1/2 and AP-2 in developmental patterning. *Nature Cell Biology*. **14**: 488-501.
59. Prosser, D.C., Drivas, T.G., Maldonado-Baéz, L., and **Wendland B.** (2011). Existence of a novel clathrin-independent endocytic pathway in yeast that depends on Rho1 and formins. *Journal of Cell Biology*. **195**:657-71. PMCID: PMC3257529. Evaluated in F1000.
58. Reider, A. and **Wendland, B.** (2011). Endocytic adaptors – social networking at the plasma membrane. *Journal of Cell Science*. **124**:1613-22. Highlighted on the cover.

57. Prosser, D., Tran, D., Schooley, A., **Wendland, B.**, and Ngsee, J.K. (2010). A novel, retromer-independent role for Sorting Nexins 1 and 2 in RhoG-dependent membrane remodeling. *Traffic*. **11**:1347-62. Evaluated in F1000.
56. Prosser, D., Whitworth, K.P. and **Wendland, B.** (2010). Quantitative analysis of endocytosis with cytoplasmic pHluorin chimeras. *Traffic*. **11**:1141-50. PMCID: PMC2919640. Evaluated in F1000.
55. Traub, L.M. and **Wendland, B.** (2010). Cell Biology: How to do a coat. *Nature*. **465**:556-7.
54. Does, M.R., Schnell, J.D., Maldonado-Báez, L., **Wendland, B.** and Hicke, L. (2010). The Function of Yeast Epsin and Ede1 Ubiquitin-Binding Domains During Receptor Internalization. *Traffic*. **11**:151-60.
53. Boettner, D.R., D'Agostino, J.L., Torres, O.T., Daugherty-Clarke, K., Uygur, A., Reider, A., **Wendland, B.**, Lemmon, S.K., and Goode, B.L. (2009). The F-BAR domain protein Syp1 negatively regulates WASp-Arp2/3 complex activity during endocytic patch formation. *Current Biology*. **19**:1979-87.
52. Reider, A., Barker, S.L., Im, Y. J., Mishra, S., Hurley, J.M., Traub, L. and **Wendland, B.** (2009). Syp1 is a conserved endocytic adaptor that contains domains involved in cargo selection and membrane tubulation. *EMBO J*. **28**:3103-16. Evaluated in F1000.
51. Mukherjee, D., Coon, B.G., Edwards III, D.F., Hanna, C.B., Longhi, S.A., McCaffery, J.M., **Wendland, B.**, Retegui, L.A., Bi, E., and Aguilar, R.C. (2009). The yeast endocytic protein Epsin-2 functions in a cell division signaling pathway. *J. Cell Sci*. **122**:2543-2464.
50. Pierce, B.D. and **Wendland, B.** (2009). Sequence of the yeast protein expression plasmid pEG(KT). *Yeast*. **26**:349-353.
49. Burston, H.E., Maldonado-Báez, L., Davey, M., Montpetit, B., Schluter, C., Wendland, B., and Conibear, E. (2009). Regulators of Yeast Endocytosis Identified by Systematic Quantitative Analysis. *J. Cell Biol*. **185**:1097-1110. Evaluated in F1000.
48. Maldonado-Báez, L., Does, M.R., Perkins, E.M., Drivas, T.G., Hicke, L., and **Wendland, B.** (2008). Interaction between Epsin/Yap180 Adaptors and the Scaffolds Ede1/Pan1 Is Required for Endocytosis. *Mol Biol Cell*. **19**:2936-2948.
47. Barker, S.L., Lee, L., Pierce, B.D., Maldonado-Báez, L., Drubin, D., and **Wendland, B.** (2007). Interaction of the endocytic scaffold protein Pan1 with the type I myosins contributes to the late stages of endocytosis. *Mol. Biol. Cell*. **18**:2893-2903.
46. McPherson, P.S., Ritter, B., and **Wendland, B.** (2009). Clathrin-mediated endocytosis. *Trafficking Inside Cells: Pathways, Mechanisms and Regulation*. Molecular Biology Intelligence Unit, Landes Bioscience, Springer Science + Business Media, LLC. Editors: N. Segev, A. Alfonso, G. Payne and J. Donaldson. 159-182.
45. Maldonado-Báez, L. and **Wendland, B.** (2006). Endocytic Adaptors: Recruiters, coordinators and regulators. *Trends in Cell Biology*. **16**: 505-513.
44. Jahren, A.H., Kelm, K., **Wendland, B.**, Petersen, G., and Seberg, O. (2006). The carbon stableisotopecomposition of dsDNA isolated from an incipient paleosol. *Geology*. **34**:381-4.
43. Aguilar, R.C., Longhi, S.A., Shaw, J.D., Yeh, L.-Y., Kim, S., Schon, A., Freire, E., Hsu, A., Watson, H.A., McCormick, W.K., and **Wendland, B.** (2006). Epsin ENTH domains perform an essential function regulating Cdc42 through binding Cdc42 GTPase activating

- proteins. *Proc Natl Acad Sci U S A*. **103**:4116-4121. [Cover highlight, and accompanying Commentary]. Evaluated in F1000.
42. Katzmann, D.J. and **Wendland, B.** (2005). Analysis of ubiquitin-dependent protein sorting within the endocytic pathway in *Saccharomyces cerevisiae*. *Methods in Enzymology*. **399**:192-211.
 41. Aguilar R.C. and **Wendland B.** (2005). Endocytosis of membrane receptors: Two pathways are better than one. *Proc Natl Acad Sci U S A*. **102**:2679-2680.
 40. Saiardi, A., Resnick, A.C., Snowman, A.M., **Wendland, B.**, and Snyder, S.H. (2005). Inositol pyrophosphates mediate cell death by regulating PI3-related protein kinases. *Proc Natl Acad Sci USA*. **102**:1911-1914.
 39. Miliaras, N.B., Park, J.-H., and **Wendland, B.** (2004). The function of the endocytic scaffold protein Pan1p depends on multiple domains. *Traffic*. **5**:963-978.
 38. Miliaras, N.B. and **Wendland, B.** (2004). EH-proteins: multivalent regulators of endocytosis (and other pathways). *Cell Biochemistry and Biophysics*. **41**:295-318.
 37. Watson, H.A., Von Zastrow, M., and **Wendland, B.** Endocytosis. (2004). In *Encyclopedia of Molecular Cell Biology and Molecular Medicine*. Second edition. Edited by Robert A. Myers. Wiley-VCH. Volume 4, 181-224.
 36. Baggett, J.J., Shaw, J.D., and **Wendland, B.** (2003). Fluorescent Labeling of Yeast. Edited by J. Lippincott-Schwartz and P. Matsudira. *Current Protocols in Cell Biology*, Unit 4.13.
 35. Baggett, J.J., D'Aquino, K.E., and **Wendland, B.** (2003). The Sla2p Talin domain plays a role in endocytosis in *Saccharomyces cerevisiae*. *Genetics*. **165**:1661-1674.
 34. Meriin, A.B., Zhang, X., Miliaras, N.B., Kazantsev, A., Chernoff, Y.O., McCaffery, J.M., **Wendland, B.**, and Sherman, M.Y. (2003). Aggregation of expanded polyglutamine domain in yeast leads to defects in endocytosis. *Mol. Cell. Biol.* **23**:7554-65.
 33. Sekiya-Kawasaki, M., Cope, M.J.T.V., Groen, A.C., Kaksonen, M., Zhang, C., Shokat, K.M., **Wendland, B.**, McDonald, K.L., McCaffery, J.M., and Drubin, D.G. (2003). Dynamic phosphoregulation of the cortical actin cytoskeleton and endocytic machinery revealed by real-time chemical genetic analysis. *J. Cell Biology*. **162**:765-72. Evaluated in F1000.
 32. Overstreet, E., Chen, X., **Wendland, B.**, and Fischer, J.A. (2003). Either portion of a severed *Drosophila* epsin (Liquid Facets) functions in the internalization of Delta in the developing eye. *Curr Biol*. **13**:854-60.
 31. Shaw, J.D., Hama, H., Sohrabi, F., DeWald, D.B., and **Wendland, B.** (2003). Phosphatidylinositol (3,5) bisphosphate is required for delivery of endocytic cargo into the multivesicular body. *Traffic* **4**:479-90.
 30. Aguilar, R.C. and **Wendland, B.** (2003). Ubiquitin: not just for proteasomes anymore. *Curr. Opin. in Cell Biol.* **15**: 184-90.
 29. Aguilar, R.C., Watson, H.A., and **Wendland, B.** (2003). The yeast epsin Ent1 is recruited to membranes through multiple independent interactions. *JBC* **278**(12):10737-43.
 28. **Wendland, B.** Epsins: adaptors in endocytosis? (2002) *Nature Rev. Mol. Cell Biol.* **3**:971-7.

27. Saiardi, A., Sciambi, C.J., McCaffery, J.M., **Wendland, B.**, and Snyder, S.H. (2002). Inositol pyrophosphates regulate endocytic trafficking. *Proc. Natl. Acad. Sci.* **99**:14206-11.
26. Hurley, J.H. and **Wendland, B.** (2002). Endocytosis: Driving membranes around the bend. *Cell.* **111**:143-6.
25. Vida, T.A. and **Wendland, B.** (2002). Flow cytometry for selection of yeast membrane trafficking mutants. Ed: C. Guthrie and G. Fink. *Methods in Enzymology* **351**: 623-631.
24. Babst, M., Katzmann, D.J., Snyder, W.B., **Wendland, B.**, and Emr, S.D. (2002). Endosome- Associated Complex, ESCRT-II, Recruits Transport Machinery for Protein Sorting at the Multivesicular Bodies. *Dev. Cell.* **3**:283-9. Evaluated in F1000.
23. Watson, H.A., Cope, M.J.T.V., Groen, A.C., Drubin, D.G. and **Wendland, B.** (2001). *In vivo* role for Actin Regulating Kinases in endocytosis and yeast epsin phosphorylation. *Mol. Biol. Cell.* **12**(11):3668-79.
22. Shaw, J.D., Cummings, K.B., Hoyer, G., Michaelis, S., and **Wendland, B.** (2001). Yeast as a model system for studying endocytosis. *Experimental Cell Research.* **271**: 1-9.
21. Duncan, M.C., Cope, M.J.T.V., Goode, B.L., **Wendland, B.**, and Drubin, D.G. (2001). Yeast Eps15-like endocytic protein, Pan1p, activates the Arp2/3 complex. *Nature Cell Biology.* **3**(7):687-90.
20. **Wendland, B.** (2001). Everything you ever wanted to know about endocytosis. Review of: *Endocytosis*, edited by M. Marsh. Volume 36 in Frontiers in Molecular Biology series, edited by B.D. Hames and D.M. Glover. Oxford University Press, 2001. *Nature Cell Biology.* **3**: E254.
19. **Wendland, B.** (2001). Round-trip ticket: Re-cycling to the plasma membrane requires RME-1. News and Views in *Nature Cell Biol.* **3**(6):E133-5.
18. Wang, G., McCaffery, J.M., **Wendland, B.**, Dupre, S., Haguenaue-Tsapais, R., and Huibregtse, J.M. (2001). Localization of the Rsp5p Ubiquitin-Protein Ligase at Multiple Sites within the Endocytic Pathway. *Mol Cell Biol.* **21**:3564-75.
17. Baggett, J.J., and **Wendland, B.** (2001). Clathrin function in yeast endocytosis. *Traffic.* **2**(5):297-302.
16. **Wendland, B.**, Steece, K.E., and Emr, S.D. (1999) Yeast Epsins contain an essential N-terminal ENTH domain, bind clathrin, and are required for endocytosis. *EMBO Journal.* **18**(16):4383-4393.
15. Kay, B.K., Yamabhai, M., **Wendland, B.**, and Emr, S.D. (1998). Identification of a novel domain shared by putative components of the endocytic and cytoskeletal machinery. *Protein Science* **8**: 435-438.
14. **Wendland, B.**, Emr, S.D., and Riezman, H. (1998). Protein traffic in the yeast endocytic and vacuolar sorting pathways. *Curr. Opin. in Cell Biol.* **10**: 513-522.
13. Babst, M., **Wendland, B.**, Estepa, E.J., and Emr, S.D. (1998). The Vps4p AAA ATPase regulates membrane association of a Vps protein complex required for normal endosome function. *EMBO J.* **17**: 2982-2993.

12. **Wendland, B.** (1998). For budding yeast investigators. Review of: *Methods in Yeast Genetics* by A. Adams, D.E. Gottschling, C.A. Kaiser, and T. Stearns. Cold Spring Harbor Laboratory Press, 1997. *Trends Cell Biol.* **8**:462-463.
11. **Wendland, B.** and Emr, S.D. (1998). Pan1p, yeast eps15, functions as a multivalent adaptor that mediates protein-protein interactions essential for endocytosis. *J. Cell Biol.* **141**: 71-84.
10. **Wendland, B.**, McCaffery, J.M., Xiao, Q. and Emr, S.D. (1996). A novel fluorescence-activated cell sorter-based screen for yeast endocytosis mutants identifies a yeast homologue of mammalian eps15. *J. Cell Biol.* **135**: 1485-1500.
9. Malgaroli, A., Ting, A.E., **Wendland, B.**, Bergamaschi, A., Villa, A., Tsien, R.W., and Scheller, R.H. (1995). Presynaptic component of long-term potentiation visualized at individual hippocampal synapses. *Science* 268, 1624-1628.
8. **Wendland, B.** and Scheller, R.H. (1994). Secretion in AtT-20 cells stably transfected with soluble synaptotagmins. *Mol. Endocrinology* 8, 1070-1082.
7. **Wendland, B.** and Scheller, R.H. (1994). Molecular mechanism of synaptic vesicle docking and membrane fusion. *Semin Neurosci* **6**: 167-76.
6. **Wendland, B.**, Schweizer, F.E., Ryan, T.A., Nakane, M., Murad, F., Scheller, R.H., and Tsien, R.W. (1994). Existence of nitric oxide synthase in rat hippocampal pyramidal cells. *Proc. Natl. Acad. Sci.* 91, 2151-2155.
5. Ryan, T.A., Reuter, H., **Wendland, B.**, Schweizer, F.E., Tsien, R.W., and Smith, S.J. (1993). The kinetics of synaptic vesicle recycling measured at single presynaptic boutons. *Neuron* 11, 713-724.
4. Miller, K.G., **Wendland, B.**, and Scheller, R.H. (1993). Identification of a 34 kd synaptic vesicle and nervous system specific protein. *Brain Res.*, 616, 99-104.
3. **Wendland, B.**, Miller, K.G., Schilling, J. and Scheller, R.H. (1991). Differential expression of the p65 gene family. *Neuron.* 6, 993-1007.
2. Ngsee J.K., Trimble W.S., Elferink L.A., **Wendland B.**, Miller K., Calakos N., Scheller R.H. (1990). Molecular analysis of proteins associated with the synaptic vesicle membrane. *Cold Spring Harb Symp Quant Biol.* **55**:111-8.
1. Ngsee, J.K., Miller, K., **Wendland, B.**, and Scheller, R.H. (1990). Multiple GTP-binding proteins from cholinergic synaptic vesicles. *J. Neuroscience.* 10, 317-322.